

BIOREMEDIATION OF DETERGENTS USING LIMNOFUNGI

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ABSTRACT

Eventhough surfactants are essentially non-toxic to man at the concentrations likely to be met in wastewaters there is a wide agreement that their presence in aquatic systems is undesirable. Hence the present study was attempted using commonly available fungi in the degradation of detergents that are commonly used in this region. With regard to *Acremonium strictum*, the degrading ability of the three detergents analysed was almost the same (Tide plus 58.92%, Surf Excel 58.33% and Challenge DP 58%). *Fusarium oxysporum*, on the other hand, recorded the maximum degrading ability with Tide (60.1%) followed Surf (58.33%) and Challenge (42.1%). *Mucor luteus*, on the other hand recorded greatest degrading ability with Tide (48.48%) followed by Challenge (39.47%) and Surf (35.89%). *Cryptococcus neoformans*, however, recorded highest degrading ability with challenge (60.52%) followed by Tide (52.63%) and Surf (48.83%) while *Aspergillus niger* recorded the highest degradability with Tide (56.25%) and equal degradability with Surf and Challenge. *Penicillium funiculum*, nevertheless recorded highest degradability with Tide (91.17%) followed by Challenge (80.1%) and Surf (74.35%). Thus, the present study shows differential degradability to different detergents by all the fungi.

Keywords: Fresh water, fungi, Bioremediation, detergents

1. INTRODUCTION

Water is an essential factor for the survival of all living organisms. Due to urbanization, chemical contamination of fresh water ecosystems has become a serious problem threatening the various life forms depending on it (BenilaSmily and Sumithra, 2016). Among the various contaminants, the use of detergents have reached alarming proportions. Detergents are formulations designed to have cleaning / solubilization properties which consist of surface-active reagents (surfactants) together with subsidiary components including builders (Stoanovicet al., 2011). As detergents have both polar and non-polar characteristics and tend to exist at phase boundaries where they are associated with both polar and non-polar media (Wemedo and Nrior, 2017). Further detergents that contain phosphate are highly caustic and surfactant detergents are toxic. Detergents including biodegradable detergents have poisonous effects in all types of aquatic life if they are present in sufficient quantities (Lenntech, 2008).

Even though surfactants are essentially non-toxic to man at the concentrations likely to be met in wastewaters there is a wide agreement that their presence in aquatic systems is undesirable (Stoanovicet al., 2011). Recent developments (Stoanovicet al., 2011; BenilaSmily and Sumithra, 2016; Wemedo and Nrior, 2017) have shown that microorganisms especially fungi have got the ability to degrade detergents. Hence the present study was attempted using commonly available fungi in the degradation of detergents that are commonly used in this region.

2. MATERIALS AND METHODS

Study Area and Sample Collection: The study area, Asur village is located in the Kumbakonam Taluk, Thanjavur District, Tamil Nadu, India. The population of the village is about 1060 and depends on Asur pond for general water resource. Fifty centimeter deep water samples were collected from Asur pond in a sterile plastic bag. The water samples collected were kept at room temperature and transferred to the laboratory and processed.

Isolation of Fungi: About 1 ml of the water sample was diluted in 99 ml of sterilized distilled water; 0.1 mL of the suspension was spread onto Petri dishes containing Potato Dextrose Agar (DA) amended with 500 mg/l chloramphenicol. The plates were allowed to incubate at 28 °C for 4 days. The colonies developed were transferred separately to PDA plates for purification. After checking the purity of the fungal colonies, they were again sub-cultured onto Potato Dextrose Agar plates.

Identification of Fungi: The macroscopic and microscopic identification of the fungi was carried out by microculture on a microscope glass slide (Raper and Thorn, 1949; Raper and Fennell, 1965; Barnett and Hunter, 1986; Subramanian, 1971; Ainsworth et al., 1973; Domschet al., 1980; Van der Plaats-Niterink, 1981; Stolk and Samson, 1983; Schipper, 1984).

Detergents: The detergents used in the present investigation were household synthetic detergent of

Challenge, Tide Plus and Surf Excel.

Detergent Degradation:The experimental fungi were separately inoculated into the flasks that contained chemically-defined microbial growth medium and the detergent to be tested. The growth medium consists of: 3 g NaNO₃, 1gK₂HPO₄, 1 g MgSO₄, 0.25 g MgSO₄.7H₂O, 0.01 gFeSO₄.7H₂O and 1 g of detergent. The flasks were incubated for 4 days. The optical density of the medium was recorded at 510 nm after four days incubation. The same setup without fungal inoculation served as control (AmiyDutt Chaturvedi and Tiwari,2013). The percentage detergent degradation was calculated using the formula:

$$\text{Percentage Degradation} = \frac{\text{Control OD} - \text{Test OD}}{\text{Control OD}} \times 100$$

3. RESULTS AND DISCUSSION

The results of the degrading ability of the various fungi collected from the systems is presented in Table-1. As evident from the table, it is clear that the fungi chosen for the present study has the ability to degrade detergents.

With regard to *Acremonium strictum*, the degrading ability of the three detergents analysed was almost the same (Tide plus 58.92%, Surf Excel 58.33% and Challenge DP 58%). *Fusarium oxysporum*, on the other hand, recorded the maximum degrading ability with Tide (60.1%) followed Surf (58.33%) and Challenge (42.1%). *Mucor luteus*, on the other hand recorded greatest degrading ability with Tide (48.48%) followed by Challenge (39.47%) and Surf (35.89%). *Cryptococcus neoformans*, however, recorded highest degrading ability with challenge (60.52%) followed by Tide (52.63%) and Surf (48.83%) while *Aspergillus niger* recorded the highest degradability with Tide (56.25%) and equal degradability with Surf and Challenge. *Penicillium funiculum*, nevertheless recorded highest degradability with Tide (91.17%) followed by Challenge (80.1%) and Surf (74.35%). Thus, the present study shows differential degradability to different detergents by all the fungi.

A comparison of the degrading ability of the present study with that of others reveals that Benila Smily and Sumithra (2016) recorded a percentage degradation of 54.29 (Surf) by *M. luteus*, 47% by *F. oxysporum*, 48% by *A. flavipes* and 42.5% by *P. piceum*. In the present study, *Fusarium* and *Penicillium* recorded higher degrading abilities while *Aspergillus* recorded almost similar degrading ability and *Mucor* recorded lower degrading ability. The differences noticed in the degradation noticed among the various detergents as well as the differential rates of degradation shown by the different fungi can be attributed to the differences in the rate of enzyme activities as well as to the differences in the composition of the detergents as opined by Benila Smily and Sumithra (2016) and Stojanovic *et al.* (2011). In addition, the variation in the physico-chemical variables and their ability to interact among themselves could also result in producing metabolic changes in these organisms (Nrior and Odokuma, 2015; Wemedo and Nrior, 2017).

In the present study, among the various fungi analysed, the highest percentage degradability was shown by *Penicillium* to all the three detergents analysed, while *Cryptococcus* recorded the second highest degradability for Challenge, *Acremonium* and *Fusarium* recorded the second highest degradability for Surf while *Fusarium* in addition also showed the second highest degradability for Tide. In the present study among all the fungi analysed, *Mucor* recorded the lowest degradability to all the three detergents.

The results of the present study clearly indicate that *P. fomicularium* which is a commonly occurring species in the aquatic system of this region can be used as a candidate species for metabolizing detergents and play an important role in the purification of aquatic systems. Table-1

Detergents powders degradation using fungal species isolated from Fresh water Pond

Species	Name of the Detergent Powder	OD of unknown sample with detergent at 510 nm	OD of known sample with detergent at 510 nm	Percentage degradation
<i>Acremonium strictum</i>	Challenge	0.62	0.36	58.00
	Tide Plus	0.56	0.33	58.92
	Surf Excel	0.72	0.42	58.33
<i>Fusarium oxysporum</i>	Challenge	0.76	0.32	42.10
	Tide Plus	0.60	0.36	60.10
	Surf Excel	0.72	0.42	58.33

Species	Name of the Detergent Powder	OD of unknown sample with detergent at 510 nm	OD of known sample with detergent at 510 nm	Percentage degradation
<i>Mucor luteus</i>	Challenge	0.80	0.30	39.47
	Tide Plus	0.66	0.32	48.48
	Surf Excel	0.78	0.26	35.89
<i>Cryptococcus neoformans</i>	Challenge	0.76	0.46	60.52
	Tide Plus	0.76	0.40	52.63
	Surf Excel	0.86	0.42	48.83
<i>Aspergillus niger</i>	Challenge	0.72	0.36	50.03
	Tide Plus	0.64	0.36	56.25
	Surf Excel	0.88	0.44	50.20
<i>Penicillium funiculum</i>	Challenge	0.75	0.60	80.00
	Tide Plus	0.68	0.62	91.17
	Surf Excel	0.78	0.58	74.35

REFERENCES

1. (IOSR-JESTET) 9:73-79. doi: 10.9790/2402-09727379
2. (IOSR-JESTET) 9:73-79. doi: 10.9790/2402-09727379
3. Abdel-Raheem, A. M. and Ali, E. H. (2004). Lignocellulolytic enzyme production by aquatic hypomyces species isolated from the Nile's delta region. *Mycopathologia*, 157: 277- 286.
4. africanus) *Journal of Environmental Sciences, Toxicology and Food Technology*
5. africanus) *Journal of Environmental Sciences, Toxicology and Food Technology*
6. Ainsworth, G. C., Sparrow, F. K. and Sussman, A. S. (1973). *Recent trends in Limnology* (Ed. Mishra, R. S.), Daya Publ. House, New Delhi, pp. 77-85.
7. AmiyDutt Chaturvedi and Tiwari, K. L. (2013). Effect of Household detergents (Surfactants) Degraded through aquatic fungi. *Recent Research in Science and Technology*, 5: 12-16.
8. Barnett, H. L. and Hunter, B. B. (1986). *Illustrated Genera of Imperfect Fungi*. 4th ed., Macmillan Publishing Company, p. 142.
9. BenilaSmily, J. M. and Sumithra, P. (2016). Degradation of household detergents using fungi isolated from fresh water ecosystem. *International Journal of Advanced Research in Biological Sciences*, 3: 120-123.
10. Domsch, K. H., Gams, W. and Anderson, T. H. (1980). *Compendium of Soil Fungi*. Academic Press, London, p. 48.
11. Lenntech, 2008. Toxicity of chemicals to aquatic species. Lenntech Water Treatment and Air Purification Holding B.V. *Journal of Aquatic, Rotterdamseweg 402 M, 2629, HH Delft, The Netherland*.
12. Nrior, R. R. and Odokuma L. O. (2015) Comparative toxicity of drilling fluids to marine
13. Nrior, R. R. and Odokuma L. O. (2015) Comparative toxicity of drilling fluids to marine water shrimp (*Mysidoposis bahia*) and brackish water shrimp (*Palaemonetes africanus*). *Journal of Environmental Sciences, Toxicology and Food Technology*, 9:73-79.
14. Nrior, R. R. and Odokuma L. O. (2015) Comparative toxicity of drilling fluids to marine
15. Raper, K. B. and Fennell, D. I. (1965). *The genus Aspergillus*, Baltimore: Williams and Wilkins Co., Baltimore. p. 686.
16. Raper, K. B. and Thorn, C. (1949). *A Manual of the Penicillia*. Williams and Wilkins, Baltimore. p. 875.
17. Schipper, M. A. A. (1984). A revision of the genus *Rhizopus* 1. The *Rhizospousstolonifera* group and *R. oryzae*. *Stud. Mycol.*, 25: 1-19.
18. Stooanovic, J., Jakovljevic, V., Maovic, I., Gajovic, O, Mijuskovic, Z. and Nedeljko, T. (2011). Influence of detergent on metabolic activity of fungi *Aspergillus niger*. *Natural Science*, 3: 466-470.
19. Stolk, A. C. and Samson, R. A. (1983). The Ascomycete genus *Eupenicillium* and related *Penicillium* anamorphs. *Stud. Mycol. Baarn.*, 23: 1-149.
20. Subramanian, C. V. (1971). The Phialide. In: *Taxonomy of Fungi imperfecti*. (Ed. B. Kendrick), Toronto. University of Toronto Press. pp. 92-115
21. Van der Plaats-Niterink, A. J. (1981). *Monograph of the Genus Pythium*. *Studies in Mycology*. No. 21, Central Bureau Voor Schimmel Culture, Baarn, The Netherlands.
22. water shrimp (*Mysidoposisbahia*) and brackish water shrimp (*Palaemonetes*
23. water shrimp (*Mysidoposisbahia*) and brackish water shrimp (*Palaemonetes*
24. Wemedo, S. A. and Nrior, R. R. (2017). Response of *Mucor racemosus* to toxicity of domestic detergents. *Journal of Emerging Trends in Engineering and Applied Sciences*, 8: 85-91.